

## RESEARCH PAPER

# A comparative study of scopolamine, D-Galactose and AlCl<sub>3</sub>-induced Alzheimer's like disease by enhancing hippocampal neurodegeneration and memory impairment in male rats

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### ABSTRACT:

Alzheimer's disease (AD) is a debilitating neurodegenerative disorder identified by progressive loss of neurons, cognition decline, and memory deficits. The current study sought out to compare the effects of scopolamine (SCO), D-galactose (D-gala), and aluminum chloride (AlCl<sub>3</sub>) on memory performance, histological alterations of the hippocampus, and to determine the best model for inducing Alzheimer's-like disease. Twenty-four male adult albino rats (250–300g) were used in this study, which were arbitrarily assigned into four groups: control group (normal saline), SCO group (2 mg/kg), D-gala group (125 mg/kg) and AlCl<sub>3</sub> group (50 mg/kg). The chemicals were given intraperitoneally for 30 days. All the rats were subjected to novel object recognition (NOR) and Barnes maze (BM) tests to assess memory performance. At the end of the period, all the rats were anesthetized, blood samples were taken for hematological and biochemical tests, and the brains were removed for morphometric analysis. The results have demonstrated that SCO, D-gala, and AlCl<sub>3</sub> caused memory impairment in BM and NOR tests, and malondialdehyde serum MDA levels significantly increased. Histological analysis showed degenerative pyknotic cells in the rat hippocampus. The impairment in memory performance and histopathological alterations were higher in the AlCl<sub>3</sub> than in the other models, but the hematological parameters were significantly changed. Therefore, it was concluded that SCO (2 mg/kg) is the best model for mimicking AD-like disease among D-gala and AlCl<sub>3</sub> by enhancing histopathological alternations and declining memory in rats without affecting other study parameters.

KEY WORDS: Alzheimer's disease, hippocampus, memory impairment, AlCl<sub>3</sub>

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### 1.INTRODUCTION :

Alzheimer's disease (AD), is the most common neurodegenerative disorder that about 46 million persons in the world suffer recently, and estimated to rise to 131.5 million by the year 2050 and progressively increase throughout the years (Mat Nuri et al., 2017). AD patients suffer from memory loss and other cognitive deficits due to cholinergic neurons degenerating or loss of neurons in the hippocampus and cortex (Kumar et al., 2018). The hippocampus is a section of the limbic system (Alsemeh et al., 2020). It has a very distinctive shape, composed of two areas: the dentate gyrus (DG) and the Cornu Ammonis (CA), which mainly contain the pyramidal cells and the granule cells, respectively.

Based on the architecture of the pyramidal neurons, the CA area can be separated into the CA1, CA2, CA3, and CA4 subareas (Boccaro et al., 2015, Brennan et al., 2019). The hippocampus is crucial for some processes involved in learning and memory (Um et al., 2017).

Many hypotheses about AD have been developed, including cholinergic neuron degeneration, oxidative stress, amyloid  $\beta$  (A $\beta$ ), tau, and inflammation (Ravi et al., 2018). Aggregation of A $\beta$  allows for neural loss and synaptic degeneration. Tau protein hyperphosphorylation hinders axonal transport from the soma to nerve terminals and vice versa (Knopman et al., 2021). The development of AD is influenced by oxidative stress. AD patients have greater levels of malondialdehyde (MDA), a byproduct of lipid peroxidation brought on by free radicals, which causes neurodegeneration that is

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facilitated by the excess creation of free radicals (Abulfadl et al., 2018).

Several kinds of AD rodent models are used to study different characteristics of the illness. Aluminum is the third most common element on the planet. It has also been suggested that an increased intake of aluminum ions may be one of the risk factors for AD and nervous system impairment (Klotz et al., 2017, Yang et al., 2019). Aluminum chloride (AlCl<sub>3</sub>) was used in many manufactured such as medicines, foods, toothpaste, and in drinking water (Newairy et al., 2009, Colomina and Peris-Sampedro, 2017). AlCl<sub>3</sub> can reach the circulation system and quickly move to the brain by passing the blood brain barrier (BBB). When AlCl<sub>3</sub> reaches the brain, it accumulates in various areas involving the hippocampus as it causes neurodegenerative disease development (Alam and Bansal, 2020, Cao et al., 2017). The toxicity of AlCl<sub>3</sub> induces a wide range of physiological, behavioral, and biochemical dysfunctions in humans and animals (Aly et al., 2018).

D-galactose is a mono-saccharide sugar, used for senescence in age-associated neurodegenerative disorders (Shwe et al., 2018, Vlassara et al., 1994). Chronic exposure of rats to D-galactose leads to gradual memory impairment and neurodegeneration (Imbimbo et al., 2005). Malondialdehyde (MDA) levels may rise resulting from the increased oxidative stress caused by D-gala exposure (Yin et al., 2010).

Scopolamine (SCO) is an anticholinergic agent that can pass BBB and then interact with the cholinergic system to impair memory and learning and produce an AD disease (Budzynska et al., 2015, Fuji et al., 2018). Cognitive impairment in AD people is strongly associated with the loss of cholinergic neurons and abnormalities in cholinergic neurotransmission in the cortex and hippocampal regions (Abd-El-Fattah et al., 2014). The objective of this study is to determine the impacts of aluminum chloride, scopolamine, and D-galactose on rat hippocampal structure and to select the best model that mimics the AD-like disease.

## 2. Materials and methods

### 2.1 Drug and Chemicals

These chemicals were used in this study: Scopolamine (Gulf Pharmaceutical Industries, Ras

Al Khaimah, U.A.E), aluminum chloride (AlCl<sub>3</sub>), and D-galactose (BDH, England).

### 2.2 Experimental Animals

Adult male rats (250–300 gram) were kept in plastic cages shielded with steel grids, six rats per cage. The environment in the room was regulated at a temperature of (23 ± 3 C<sup>o</sup>), with a 12-hour cycle for light and dark. During the experiment, drinking, water, and a standard food were provided to each cage *ad libitum*. Before the start of the experiment, all the rats were acclimatized for 2 weeks. Only the smallest number of rats was required to create reliable logical data. Any stress and pain during the experiment were reduced.

### 2.3 Experimental Design

The idea was to use intraperitoneal injection (I.P) with of scopolamine (SCO), aluminum chloride (AlCl<sub>3</sub>), or D-galactose (D-gala) so as to induce memory impairment in rats and to evaluate behavioral, biochemical, and histological effects on rats. Twenty four adult male Wistar rats were arranged arbitrarily into 4 groups containing six animals each: the control group received normal saline intraperitoneally, scopolamine group received scopolamine at a dose of 2 mg/kg (Hafez et al., 2017). The AlCl<sub>3</sub> group received aluminum chloride (50 mg/kg body weight), which was melted in normal saline (Chavali et al., 2020). The D-galactose group was injected with D-gala at a dose of 125 mg/kg body weight, which was melted in normal saline (Li et al., 2020). The treatments were carried out for 30 days.

### 2.4 Body weight evaluation

Rat body weight was recorded on the first day and the last day of the experiment.

### 2.5 Behavioral experiments

The behavioral tests were performed after the end period of treatment to evaluate the effects of D-gala, SCO, and AlCl<sub>3</sub> on memory performance by novel object recognition and the Barnez maze tests. A camera (C270 HD Webcam Logitech) was used to record the activity of animals.

#### 2.5.1 Novel object recognition test (NOR)

Hippocampal function and memory recognition, especially spatial and working memory in rats, were evaluated by using the NOR test (Singh et al., 2018). This test was carried out in an open box (45 × 40 × 46 cm). The NOR test

was done (9:30 AM to 5:30 PM) under red light exposure (40 LX) (Retinasamy et al., 2019). In order to habituate the animals, they were allowed to explore the box for ten minutes. After 24 hours, the rat was placed inside the open box between two identical objects (familiar) and was permitted to explore the familiar objects for 4 minutes. Then, the rat was removed from the box and transported into their cage, and the box was cleaned with 60% ethanol to eliminate the effect of odor on rat behavior. After 20 minutes, the rat was exposed to a new object (novel) in addition to the familiar object for three minutes (Justin-Thenmozhi et al., 2018). The discrimination index (DI) was defined as the percentage of time spent on a new object to the total time spent on familiar and novel objects together (Batoool et al., 2016).

### 2.5.2 Barnes Maze Test (BM)

The Barnes maze is a broadly accepted test of hippocampal-dependent learning and memory in rodents (Zhang et al., 2018). In brief, the current test was performed on a white circular stage (126 cm in diameter) with 18 holes (10 cm in diameter), 4 cm away from the edge, and no surrounding walls. The maze was raised 90 cm from the base and light intensity (1250 LX) was determined by using a digital lux meter. A light-emitting diode (LED) light source was installed on the ceiling nearly 120 cm from the top of the maze to increase rats' motivation to escape from the circular platform and to search for the target hole. A small removable black box (escape box) was put at the button of one hole (target hole) in which the animal could hide. The BM test involves the following stages: habituation (1 day), acquisition stage (five days), and probe phase (1 day). The maze and escape box were cleaned with 60% alcohol after each rat was tested. Based on the techniques previously used by other researchers, the experimental design was developed (Sahraei et al., 2019). Latency time and time spent to reach the target zone were documented during the acquisition phase and the probe phase, respectively.

### 2.6 Organs and Blood Samples Collection

After BM and NOR tests, rats were fasted for 13 hours and then anesthetized with an intraperitoneal (I.P) injection of ketamine (90 mg/kg) and xylazine (12 mg/kg) (Farzampour et al., 2016). Blood was collected via the left ventricle. Then it was separated into two parts. The first part was collected into centrifuge tubes

without anticoagulant. The blood was centrifugated at 2400 rpm for 10 minutes, and then the serum samples were obtained, which were used for the determination of the MDA and serum electrolyte. The second portion was collected into sterilized tubes containing dipotassium salts of ethylenediaminetetraacetic acid (EDTA) for hematological study. Each brain was swiftly removed after it was sacrificed by decapitation.

### 2.7 Blood glucose determination

Control and experimental rats were fasted for 13 hours but provided with tap water *ad libitum*. Blood samples from the rats were collected by the heart puncture method. Blood glucose was measured by using a blood glucose monitor (VivaChek Laboratories, Wilmington, USA).

### 2.8 Hematological analysis

An automated blood cell analyzer (Boule Medical AB, Stockholm, Sweden) was used to determine platelet distribution width (PDW), red blood cells (RBCs), white blood cells (WBCs), and hemoglobin (HGB).

### 2.9 Determination of serum electrolytes

Serum electrolytes ( $\text{Ca}^{2+}$ ,  $\text{Na}^+$ ,  $\text{K}^+$  and  $\text{Cl}^-$ ) were determined from different groups by using the Centromeric GmbH Wartenberg, Germany reagent kit.

### 2.10 Serum malondialdehyde (MDA) determination

Malondialdehyde (MDA) was measured according to this method (Ohkawa et al., 1979). The principle is the reaction between the MDA in the serum and thiobarbituric acid (TBA). The optical density was scanned spectrophotometrically (SQU10111012002, USA) at 535 nm. The optical density was changed to concentration ( $\mu\text{mol/L}$ ).

### 2.11 Histological section preparation

At the end of the period, animals were killed, and the whole brains of all the animals were swiftly removed. The brains were directly fixed in formalin (10%) for 48 hours, washed and dehydrated with graded ethanol, cleared in xylol, then set in wax. Paraffin blocks were cut at 4- $\mu\text{m}$  thickness via a rotary microtome (SRM 200 CW, Netherland) and dyed with haematoxylin and eosin (H&E) (Tamizhazhagan and Pugazhendy, 2017) for studying the histological structure of the

rat hippocampus by using a light microscope (Olympus, 810812, Japan).

### 2.12 Statistical analysis

To perform the statistical analysis, GraphPad Prism (Version 8.0.1) was used. Kolmogorov-Smirnov and Shapiro-Wilk tests were used for all data to check normality tests. All data are represented as mean  $\pm$  SEM. One-way and or two-way ANOVA was performed for comparison, followed by a Tukey post hoc test. A difference is considered statistically significant if  $p \leq 0.05$ .

## 3. Results

### 3.1 Effect of SCO, D-gal and AlCl<sub>3</sub> on body weight, blood glucose, and hematological parameters

**Table 1** Effect of SCO, D-gal and AlCl<sub>3</sub> on body weight, blood glucose, and hematological parameters.

| Parameter                       | N | Control                        | SCO                            | D-gala                         | AlCl <sub>3</sub>               |
|---------------------------------|---|--------------------------------|--------------------------------|--------------------------------|---------------------------------|
| Initial weight (g)              | 6 | 267.5 $\pm$ 3.09 <sup>a</sup>  | 265 $\pm$ 5.24 <sup>a</sup>    | 271 $\pm$ 6.09 <sup>a</sup>    | 281.7 $\pm$ 4.59 <sup>a</sup>   |
| Final weight (g)                | 6 | 332.2 $\pm$ 7.23 <sup>a</sup>  | 335.8 $\pm$ 2.93 <sup>a</sup>  | 337.2 $\pm$ 17.28 <sup>a</sup> | 253.2 $\pm$ 7.41 <sup>b</sup>   |
| Body weight gain (g)            | 6 | 76.67 $\pm$ 9.43 <sup>a</sup>  | 71.67 $\pm$ 3.50 <sup>a</sup>  | 58.33 $\pm$ 6.80 <sup>a</sup>  | -45.17 $\pm$ 15.34 <sup>b</sup> |
| Blood glucose (mg/dl)           | 6 | 110.5 $\pm$ 10.18 <sup>a</sup> | 113.2 $\pm$ 5.64 <sup>a</sup>  | 120.2 $\pm$ 4.57 <sup>a</sup>  | 121.3 $\pm$ 7.23 <sup>a</sup>   |
| RBC (10 <sup>6</sup> / $\mu$ L) | 6 | 7.745 $\pm$ 0.18 <sup>a</sup>  | 6.922 $\pm$ 0.36 <sup>a</sup>  | 7.646 $\pm$ 0.06 <sup>a</sup>  | 5.979 $\pm$ 0.22 <sup>b</sup>   |
| HGB (g/dL)                      | 6 | 14.957 $\pm$ 0.45 <sup>a</sup> | 13.18 $\pm$ 1.10 <sup>a</sup>  | 13.92 $\pm$ 0.70 <sup>a</sup>  | 11.77 $\pm$ 0.26 <sup>b</sup>   |
| WBC (10 <sup>3</sup> / $\mu$ L) | 6 | 6.040 $\pm$ 1.13 <sup>a</sup>  | 7.240 $\pm$ 0.47 <sup>ab</sup> | 6.860 $\pm$ 0.33 <sup>ab</sup> | 9.218 $\pm$ 0.78 <sup>b</sup>   |
| PDW (fl )                       | 6 | 8.733 $\pm$ 0.15 <sup>a</sup>  | 11.50 $\pm$ 4.62 <sup>a</sup>  | 8.833 $\pm$ 0.25 <sup>a</sup>  | 19.65 $\pm$ 5.16 <sup>b</sup>   |

Values are represented as mean  $\pm$  SEM (n=6). Different alphabetical letters (a and b) represent significant difference among the groups. One-way ANOVA with Tukey's post hoc test was applied,  $p < 0.05$ . RBC: red blood cells, HGB: hemoglobin, WBC: white blood cells, PDW: platelet distribution width, SCO: scopolamine, D-gala: D-galactose and AlCl<sub>3</sub> aluminum chloride.

### 3.2 Effect of SCO, D-gala and AlCl<sub>3</sub> on serum electrolytes and malondialdehyde (MDA)

The results of the mean values of serum electrolytes (Ca<sup>2+</sup>, Na<sup>+</sup>, K<sup>+</sup> and Cl<sup>-</sup>) and MDA are shown in Table 2. Statistical analysis in this present study showed that serum Ca<sup>2+</sup> in the AlCl<sub>3</sub> treated group significantly decreased (\* $p < 0.05$ ) in the concentration compared to control group. In D-gala group, there is a significant increase (\* $p < 0.05$ ) in the serum Cl<sup>-</sup> than SCO, AlCl<sub>3</sub> and

The results of rat body weight, blood glucose levels, and hematological parameters in the SCO, D-gal, and AlCl<sub>3</sub> groups are shown in Table 1. The results revealed that, administration of AlCl<sub>3</sub> to the experimental animals significantly reduced (\*\* $p < 0.001$ ) in the RBC, HGB, and body weight in comparison with other groups. The WBC count in the SCO treated groups increased but not significantly, while in the AlCl<sub>3</sub> treated group, WBC count and PDW significantly increased when compared to WBC count and PDW of other treatment groups. The automated blood cell analyzer could not measure the platelet count except for one sample, which was 1159  $\times 10^3/\mu$ L, due to high count level. Blood glucose did not change significantly among groups.

control groups. In addition, a significant reduction (\*\* $p < 0.001$ ) in the concentration of serum Na<sup>+</sup> was detected in the D-gala when compared to the SCO, AlCl<sub>3</sub>, and control groups. However, serum K<sup>+</sup> did not show a significant change among groups. MDA levels in serum were significantly increased in SCO (\*\* $p < 0.001$ ), D-gala (\* $p < 0.05$ ) and AlCl<sub>3</sub> (\*\* $p < 0.001$ ) groups than control group.



**Table 2** Effect of SCO, D-gal and AlCl<sub>3</sub> on serum electrolytes (Ca<sup>2+</sup>, Na<sup>+</sup>, K<sup>+</sup> and Cl<sup>-</sup>) and MDA

| Parameter                | N | Control                  | SCO                       | D-gala                    | AlCl <sub>3</sub>         |
|--------------------------|---|--------------------------|---------------------------|---------------------------|---------------------------|
| Ca <sup>2+</sup> (mg/dl) | 6 | 10.07±0.217 <sup>a</sup> | 9.543±0.91 <sup>ab</sup>  | 9.497±143 <sup>ab</sup>   | 9.258±0.255 <sup>b</sup>  |
| Na <sup>+</sup> (mmol/L) | 6 | 162.5±2.74 <sup>a</sup>  | 155.9±3.57 <sup>a</sup>   | 136.2±2.58 <sup>b</sup>   | 157±1.65 <sup>a</sup>     |
| K <sup>+</sup> (mmol/L)  | 6 | 3.022±0.223 <sup>a</sup> | 3.230±0.193 <sup>a</sup>  | 3.428±0.093 <sup>a</sup>  | 3.460±0.187 <sup>a</sup>  |
| Cl <sup>-</sup> (mmol/L) | 6 | 96.32±0.812 <sup>a</sup> | 96.72±0.745 <sup>ab</sup> | 104.2±2.750 <sup>cd</sup> | 97.94±1.414 <sup>ad</sup> |
| MDA (µmol/L)             | 6 | 4.0±0.447 <sup>a</sup>   | 8.66±0.557 <sup>b</sup>   | 6.83±0.477 <sup>b</sup>   | 10±0.816 <sup>b</sup>     |

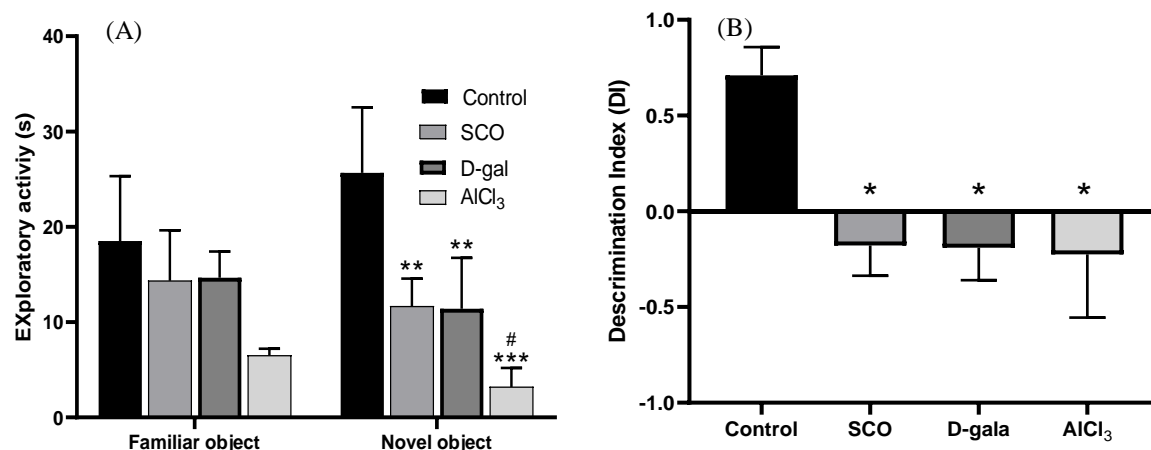
Values are represented as mean ± SEM (n=6). Different alphabetical letters (a, b, c, and d) represent significant difference among the groups. One-way ANOVA with Tukey's post hoc test was applied,  $p < 0.05$ . SCO: scopolamine, D-gala: D-galactose, AlCl<sub>3</sub>: aluminum chloride, MDA: malondialdehyde.

### 3.3 Behavioural experiments

#### 3.3.1 Effects of SCO, D-gala and AlCl<sub>3</sub> on recognition memory impairment using NOR Test

Recognition memory was detected by NOR test in rats. In our results, statistical analysis showed a significantly decreased time spent with the new object in SCO (\*\* $p < 0.01$ ), D-gala (\*\* $p < 0.01$ ) and AlCl<sub>3</sub> (\*\*\*) ( $p < 0.001$ ) groups in

comparison to control group. This is indicated by the alteration of memory recognition. Also, there was a significant difference (<sup>#</sup> $p < 0.05$ ) in AlCl<sub>3</sub> compared to SCO, and D-gala groups (Figure 1A). The discrimination index (DI) was significantly lower in SCO (\* $p < 0.05$ ), D-gala (\* $p < 0.05$ ) and AlCl<sub>3</sub> (\* $p < 0.05$ ) groups in rats than control group (Figure 1B). All results proved that SCO, D-gala, and AlCl<sub>3</sub> induced loss of recognition memory.



**Figure 1:** Effect of SCO (2 mg/kg), D-gal (125 mg/kg) and AlCl<sub>3</sub> (50 mg/kg) groups on working memory. (A) Exploratory activity. Animals in the SCO, D-gala, and AlCl<sub>3</sub> groups spent less time with novel object than familiar object when compared with control group. (B) discrimination index decreased in the SCO, D-gala, and AlCl<sub>3</sub> when compared with control group. Data are represented as means ± SEM (n=6). Statistical analysis was done by one-way ANOVA for discrimination index and two-way ANOVA for exploratory activity, followed by Tukey's Multiple Comparisons test. \*\*\* $p < 0.001$ , \*\* $p < 0.01$ , \* $p < 0.05$ : statistically different than control group. # $p < 0.05$  statistically different than SCO and D-gala groups. SCO: scopolamine, D-gala: D-galactose and AlCl<sub>3</sub>: aluminum chloride.

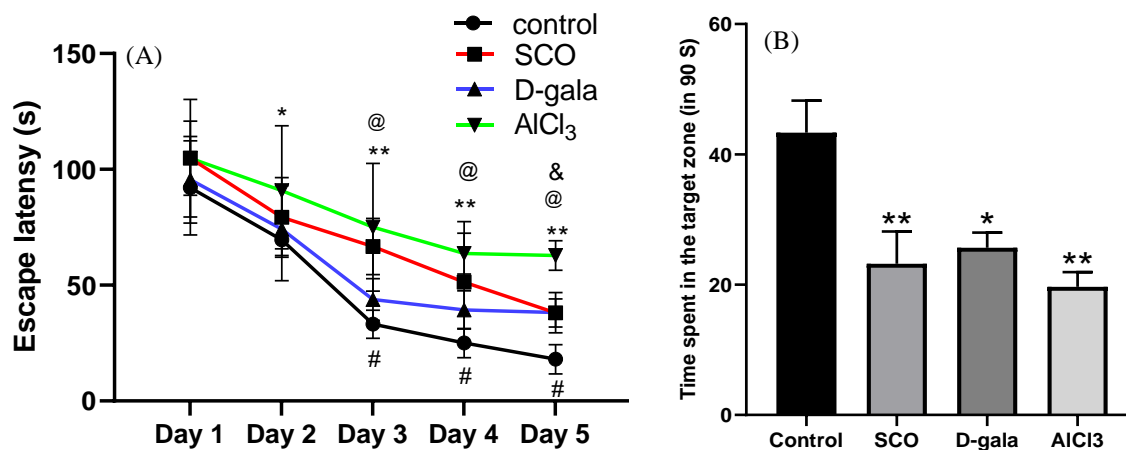
### 3.3.2 Effects of SCO, D-gala and AlCl<sub>3</sub> on long term memory impairment using BM Test

The Barnes maze (BM) is a well-known test of learning and memory in rats that depends on the hippocampus region. Learning and memory

were evaluated by the measure of escape latency to find the escape box in the acquisition phase (days 1-5) and time spent in the target zone, which before had the escape box in the probe phase (day 7). Statistical analysis revealed no significant variation in the escape latency between treatment groups on the day one of the acquisition phase. The post-hoc analysis demonstrated a significant increase (\**p* < 0.05) in the latency time (the time to reach the escape box was greater) in the rats treated with AlCl<sub>3</sub> was noted on the

second day of the acquisition phase than control group. However, there was no significant variation among SCO and D-gala than control group. On the third and fourth days of the acquisition phase, rats in SCO (\**p* < 0.05) and AlCl<sub>3</sub> (\*\**p* < 0.01) treated groups significantly need more time to find the escape box than control group. In contrast, there was no significant variation in the escape latency of the D-gala treated group (Figure 2A).

The last day of the BM test is considered as a probe phase. In this phase, escape box was removed. The post-hoc analysis indicated on day seven of BM test, time spent in the target zone in SCO (\*\**p* < 0.01), D-gala (\**p* < 0.05) and AlCl<sub>3</sub> (\*\**p* < 0.01) groups significantly decreased when compared to the control group (Figure 2B).

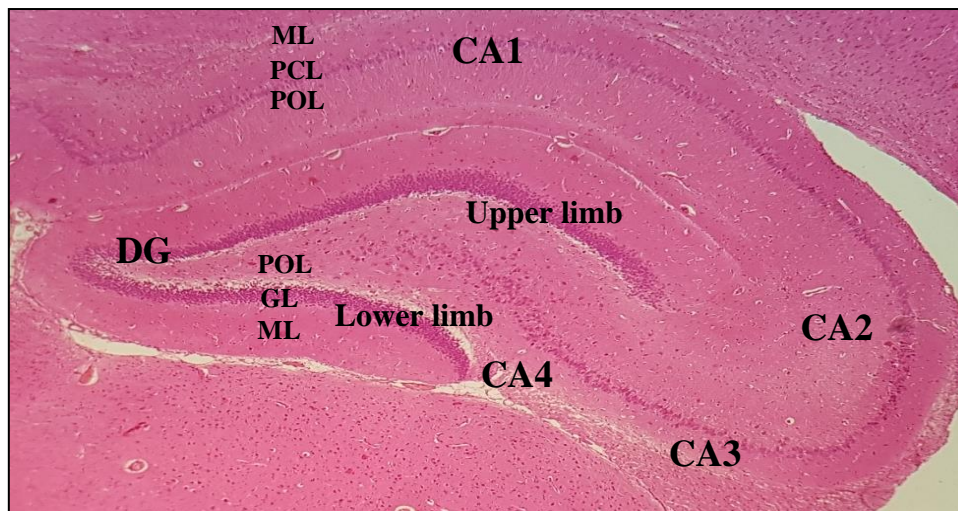


**Figure 2:** Impairment of learning and memory performance by SCO (2 mg/kg), D-gala (125 mg/kg), and AlCl<sub>3</sub> (50 mg/kg) in Barnes maze test. (A) Escape latency to find the target hole which contained an escape box on the first day of acquisition phase to find the escape box no significant difference between groups. On the second day of acquisition phase, no significant difference was found between groups. However, significant variation was only found between control and AlCl<sub>3</sub> groups (\**p* < 0.05). On the third and fourth day of acquisition phase rats in SCO and AlCl<sub>3</sub> treated groups significantly (@*p* < 0.05 and \*\**p* < 0.01) respectively need more time to find the escape box. Also, there was a significant difference (#*p* < 0.05) between D-gala, and AlCl<sub>3</sub> groups. On the fifth day of the acquisition phase, rats in SCO, D-gala, and AlCl<sub>3</sub> treated groups significantly need more time to find the escape box. Also, AlCl<sub>3</sub> differ (#*p* < 0.05) from SCO and D-gala groups. (B) Time spent in the target zone. During the probe phase on the seventh day of the Barnes maze test, rats in SCO, D-gala, and AlCl<sub>3</sub> groups (@*p* < 0.05, &*p* < 0.05 and \*\**p* < 0.01) respectively spent significantly less time in the target zone, which contained escape box. In each group data are represented as means ± SEM (n=6). Statistical analysis was done by two-way ANOVA for escape latency and one-way ANOVA for time spent in the target zone, followed by Tukey's Multiple Comparisons test. \**p* < 0.05; \*\**p* < 0.01 significantly difference compared to control group, @ *p* < 0.05 differ significantly in comparison with SCO and D-gala groups.

### 3.5 Histological Observations

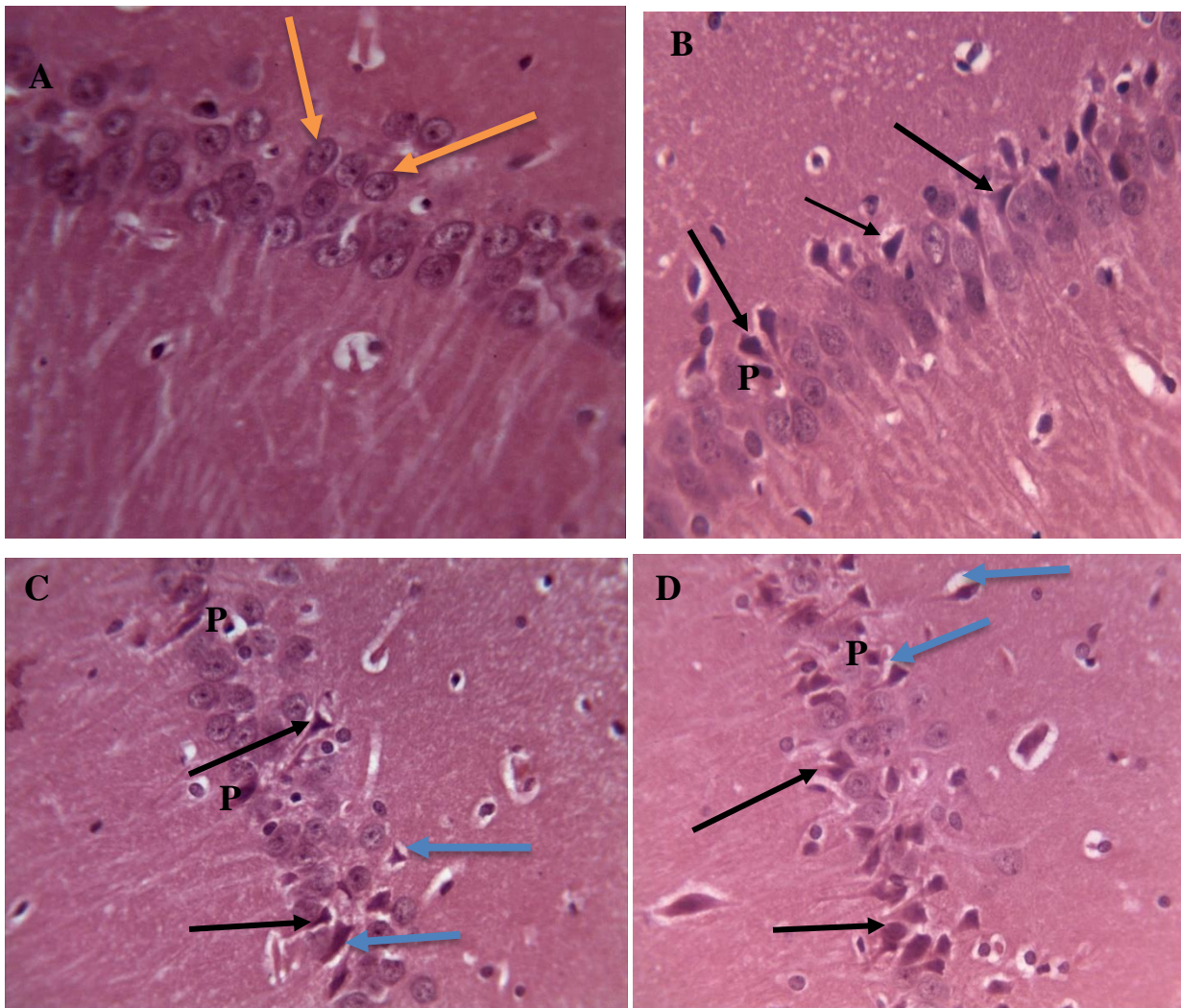
Microscopical examination of coronal sections of rats' hippocampal tissues from control, SCO, D-gala, and AlCl<sub>3</sub> groups showed the same histological features. H&E staining of the hippocampus revealed it was composed of two regions, hippocampus proprius and the dentate gyrus (DG). The hippocampus proprius can be divided into 4 Cornu Ammonis regions. These regions (CA1, CA2, CA3 and CA4) consist of molecular layer (ML), pyramidal layer (PCL), polymorphic cell layer (POL) and the dentate gyrus appeared as a coiled structure with upper and lower limbs were directed to the hippocampus

proprius. The DG composed of 3 layers, polymorphic cell layer (POL), granular cell layer (GCL), and molecular cell layer (ML) (Figure 3). The pyramidal layer of the CA1 area in the control group contained small pyramidal cells with round vesicular nuclei, visible nucleoli (Figure 4: A). In the current study, histological analysis of the pyramidal cell layer of CA1 region in the SCO, D-gala, and AlCl<sub>3</sub> groups showed pyramidal cells with shrunken degenerated nuclei and darkly pyknotic nuclei and vaculation around cells (Figure 4: B, C, D). However, more neural cells damage in AlCl<sub>3</sub> than in other models.



**Figure 3:** A coronal section of adult rat hippocampus of the control group shows dentate gyrus (DG) and hippocampus proper stained with H&E (10X). The hippocampus proper, which consists of the Cornu Ammonis (CA4, CA3, CA2, and CA1). ML: molecular layer, PCL; pyramidal cell, POL; polymorphic cell layer, GL; granular cell layer.





**Figure 4:** Section of rat hippocampus brain showing CA1 region treated with (A) normal saline (B) scopolamine (2 mg/kg), (C) D-galactose (125 mg/kg) and (D) aluminum chloride ( $\text{AlCl}_3$ ) (50 mg/kg) treated groups intraperitoneally injected for 30 days. In the control group, showed healthy neurons (normal pyramidal cells) with round vesicular nuclei and visible nucleoli (green arrows). In scopolamine, D-galactose and aluminum chloride treated groups, pyramidal cells with shrunken elongated nuclei (black arrows), vacuolation around them (blue arrows) or pyknotic nuclei (P). Stained with Hematoxylin-Eosin dye (H&E, 40 x).

#### 4. Discussion

Alzheimer's disease (AD), is the most common neurodegenerative disease (Karran and De Strooper, 2016). It is mainly categorized by the loss of cognitive and memory functions (Livingston et al., 2020) as a result of the loss of pyramidal neurons and cholinergic neurons in the brain cortex and hippocampus being degenerated (Hansson et al., 2006). The hippocampus is a portion of the brain in humans and vertebrates. It plays a vital role in memory formation (Knierim, 2015, Saeed et al., 2021). In this study, to induce memory disorders in rodents, and to select the best

model that mimics the AD-like symptoms, the SCO, D-gala, and  $\text{AlCl}_3$  models were used. This work, revealed noticeable impairments in learning and memory along with damaged neurons in the hippocampus regions in rats. In the present study, the tests for memory assessment and learning related to hippocampus were used, including Barnes maze (BM) as well as novel object recognition (NOR). These tests can assess animals' cognitive impairment (Kim et al., 2014).

The NOR test in our study showed injections of SCO, D-gala, and  $\text{AlCl}_3$  in rats caused a significant reduction in the time need for exploring the novel (new) object by measuring



time spent between novel and old objects when we compared it to the familiar object. However, the  $\text{AlCl}_3$  group is statistically more significant in comparison to the SCO and D-gala groups. Also, the NOR test revealed a significant decrease in discrimination index in the SCO, D-gala, and  $\text{AlCl}_3$  treated groups. This current study suggests that prolonging SCO, D-gala, and  $\text{AlCl}_3$  administration led to recognition memory impairment, which is also confirmed by other studies (Wong-Guerra *et al.*, 2017, Alghamdi, 2018, Saenno *et al.*, 2022).

Barnes maze (BM) test is used for measuring spatial learning and memory in animals. The first phase of the test refers to the acquisition phase, which allows evaluation of spatial learning, while the second phase of the test refers to the probe phase to evaluate spatial memory. These phases are thought to be related to hippocampus function (Barnes, 1979, Gawel *et al.*, 2019). In the current work, the BM test revealed that injection of SCO, D-gala, and  $\text{AlCl}_3$  in rat models had effects on memory and learning capability by measuring time spent in the target zone and escape latency. In this test, we noticed that SCO, D-gala, and  $\text{AlCl}_3$ -treated groups spent more time to find the escape box location in the acquisition phase. Additionally, decrease spent time in the target zone than control rats in the probe phase (Fuji *et al.*, 2018, F Ewida and A Mansour, 2015). However,  $\text{AlCl}_3$  was more effective than other models.

The histopathology characteristics of AD include the continuous degeneration of specific brain neurons (Ravi *et al.*, 2018, Chiroma *et al.*, 2019). The first area affected by this degenerative process in AD patients is the hippocampus. It plays a vital function in memory processes (Padurariu *et al.*, 2012, Heo *et al.*, 2014). The hippocampus is highly susceptible to neuronal damage. In the recent study, histopathological analysis of the brain tissues from the SCO, D-gala, and  $\text{AlCl}_3$  treated groups revealed major morphological changes and degenerate neurons in the hippocampal CA1 regions, pyramidal cells with shrunken degenerated nuclei, darkly pyknotic nuclei, and vacuolation around cells in the CA1 hippocampal region compared to the control group. These results are in accordance with those of (Fawzi *et al.*, 2020, Aboelwafa *et al.*, 2020, Firdaus *et al.*, 2022). However,  $\text{AlCl}_3$  causes more neural cell damage compared to the SCO, and D-

gala models. SCO is related to cholinergic dysfunction and oxidative stress in the hippocampus. (Pattanashetti *et al.*, 2017). In the body, d-galactokinase and galactose-one-phosphate uridyl transferase metabolize the reducing sugar D-gala. Yet high levels of D-gala result in abnormal metabolism. The conversion of D-gala to galactitol, which is not processed but aggregated in the cells, made osmotic stress and the creation of oxidative stress in the cell, which causes neurodegeneration and causes behavioral disorders and cognitive disorders (Kumar *et al.*, 2011, Wang *et al.*, 2020).  $\text{AlCl}_3$  can induce neurological conditions for instance AD (Maya *et al.*, 2016, Inan-Eroglu and Ayaz, 2018). The mechanism of  $\text{AlCl}_3$ -induced neurodegeneration is not obviously known. However,  $\text{AlCl}_3$  increases the activity of ferrous ( $\text{Fe}^{2+}$ ) and ferric ( $\text{Fe}^{3+}$ ) ions as a reason to oxidative damage, leading to neurodegeneration, which is also confirmed by another study (Aljarari and Bawazir, 2019).

In the current study, body weight was elevated in the control, SCO, and D-gala groups with age increasing, but in the groups treated with  $\text{AlCl}_3$  it was decreased significantly. These results we obtained indicate the impact of  $\text{AlCl}_3$  on body weight but that the effects of SCO and D-gal on this parameter were not significant. The underlying cause of the effect  $\text{AlCl}_3$  on body weight might be due the damage in pancreatic tissue and reduced glucose transporter 4 (GLUT4) expression level, which in turn results in glucose metabolism change (Wei *et al.*, 2018). The glucose level in the blood of the groups did not show a significant difference (Harandi *et al.*, 2015, Nam *et al.*, 2016).

One of the factors has an effect on pathophysiology of AD is oxidative stress (Ferreira *et al.*, 2015). AD patients have higher levels of MDA, which is a free radical byproduct of lipid peroxidation reaction. MDA causes cross-linking and polymerization of large molecules like nucleic acids and proteins, and has certain cytotoxicity (Xu *et al.*, 2019). In the recent study, serum MDA levels were significantly increased in the SCO, D-gala, and  $\text{AlCl}_3$  groups in comparison to the control group (Ghanbari *et al.*, 2019, Kou *et al.*, 2017, KANDIŞ *et al.*, 2022). However, in the  $\text{AlCl}_3$  group it was statistically highly significant.

According to hematological results,  $\text{AlCl}_3$  led to a significantly decreased RBC and HGB compare to control group. Also, administration of

SCO, and D-gala led to decreased RBC and HGB but no significantly lower than control group. By decreasing  $\text{Fe}^{2+}$  binding to ferritin and disrupting heme production,  $\text{AlCl}_3$  can alter membrane structures and enzyme activity for RBC production. The reduction in HGB content might be due to an increased rate of destruction or decrease in the rate of development of RBCs (Aziz and Zabut, 2011, Adedosu et al., 2018). Also, reductions in RBC and HGB might be attributed to hyper-activity of the bone marrow, leading to synthesis of RBCs with decreased integrity that are easily destroyed in the blood circulation. The reduction of RBC, HGB by  $\text{AlCl}_3$  in this study suggests the induction of anemia. Also, hematological results indicated that  $\text{AlCl}_3$  led to a significantly increased WBC when compared to SCO, D-gala, and control groups (Yakubu et al., 2017). Increase in WBC is suggestive of leukocytosis and anemia as seen from the corresponding decreases in RBC which may indicate the activation of the immune system to form defensive cells (Adedosu et al., 2018).

Electrolytes have a significant role in the maintenance of the function of muscle and nerve and also in the hydration process of the body. In this work,  $\text{AlCl}_3$  produced a significant reduction in the serum concentration of  $\text{Ca}^{2+}$  but the serum levels of  $\text{Na}^+$ ,  $\text{K}^+$ , and  $\text{Cl}^-$ , did not change significantly. D-gala produced a significant decrease in the serum concentration of  $\text{Na}^+$  and a significant rise in the serum concentration of  $\text{Cl}^-$ . On the other hand, SCO did not affect serum electrolytes (Ugbaja et al., 2017, Ogunlade et al., 2022).

## 5. Conclusion

The present study indicated that administration of SCO, D-gala, and  $\text{AlCl}_3$  induced learning and memory impairment in the Barnes Maze and novel object recognition tests by increasing the escape latency to find the escape box, decreasing the time spent in the target zone in the BM test, and lowering the recognition index in the NOR test. Additionally, histopathology results showed an increase number of shrinking or degenerating pyramidal cells in the rat hippocampus. We concluded that SCO (2 mg/kg) is the best model for mimicking AD-like disease among D-gala, and  $\text{AlCl}_3$  by enhancing histopathological alternation and impairing

memory in rats without changing hematological and electrolyte parameters.

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## Conflicts of Interest

No conflict of interest

## References

- ABD-EL-FATTAH, M. A., ABDELAKADER, N. F. & ZAKI, H. F. 2014. Pyrrolidine dithiocarbamate protects against scopolamine-induced cognitive impairment in rats. *Eur J Pharmacol*, 723, 330-8.
- ABOELWAFI, H. R., EL-KOTT, A. F., ABD-ELLA, E. M. & YOUSEF, H. N. 2020. The Possible Neuroprotective Effect of Silymarin against Aluminum Chloride-Prompted Alzheimer's-Like Disease in Rats. *Brain Sci*, 10.
- ABULFADL, Y. S., EL-MARAGHY, N. N., AHMED, A. A. E., NOFAL, S. & BADARY, O. A. 2018. Protective effects of thymoquinone on D-galactose and aluminum chloride induced neurotoxicity in rats: biochemical, histological and behavioral changes. *Neurol Res*, 40, 324-333.
- ADEDOSU, O., ADELEKE, G., ALAO, T., OJUGO, N. & AKINSOJI, O. 2018. Effects of vitamin E administration on certain biochemical and antioxidants indices in some rat tissues treated with aluminium chloride. *Annual Research & Review in Biology*, 1-13.
- ALAM, M. A. & BANSAL, G. 2020. Effects of Insulin against Aluminium Induced Neurotoxicity in Wistar Rats. *Pharmacology and Clinical Pharmacy Research*, 5, 62-71.
- ALGHAMDI, B. S. A. 2018. Possible prophylactic anti-excitotoxic and anti-oxidant effects of virgin coconut oil on aluminium chloride-induced Alzheimer's in rat models. *J Integr Neurosci*, 17, 593-607.
- ALJARARI, R. & BAWAZIR, A. 2019. Effect of Black Raisins (*Vitis vinifera*) on Aluminum Chloride Induced Alzheimer's Disease in Male Albino Rat. *International Journal of Pharmaceutical Research & Allied Sciences*, 8.
- ALSEMEH, A. E., IBRAHIM, A. A. S. & MOHAMMED, H. O. 2020. Structural and Behavioural Changes in Rat Hippocampus Induced by Methotrexate and The potential ameliorative Effect of Alpha lipoic acid. *Egyptian Journal of Histology*, 43, 301-324.
- ALY, H., ELRIGAL, N., ALI, S., RIZK, M. & EBRAHIM, N. 2018. Modulatory effects of *Casimiroa edulis* on

- aluminium nanoparticles-associated neurotoxicity in a rat model of induced Alzheimer's disease. *J. Mater. Environ. Sci*, 9, 1931-1941.
- AZIZ, I. A. & ZABUT, B. M. 2011. Determination of blood indices of albino rats treated with aluminum chloride and investigation of antioxidant effects of vitamin E and C. *Egyptian Journal of Biology*, 13, 1-7.
- BARNES, C. A. 1979. Memory deficits associated with senescence: a neurophysiological and behavioral study in the rat. *J Comp Physiol Psychol*, 93, 74-104.
- BATOOL, Z., SADIR, S., LIAQUAT, L., TABASSUM, S., MADIHA, S., RAFIQ, S., TARIQ, S., BATOOL, T. S., SALEEM, S., NAQVI, F., PERVEEN, T. & HAIDER, S. 2016. Repeated administration of almonds increases brain acetylcholine levels and enhances memory function in healthy rats while attenuates memory deficits in animal model of amnesia. *Brain Res Bull*, 120, 63-74.
- BOCCARA, C. N., KJONIGSEN, L. J., HAMMER, I. M., BJAALIE, J. G., LEERGAARD, T. B. & WITTER, M. P. 2015. A three-plane architectonic atlas of the rat hippocampal region. *Hippocampus*, 25, 838-57.
- BRENNAN, P. P., STANDRING, S. & WISEMAN, S. 2019. *Gray's surgical anatomy e-book*, Elsevier Health Sciences.
- BUDZYNSKA, B., BOGUSZEWSKA-CZUBARA, A., KRUK-SLOMKA, M., SKALICKA-WOZNIAK, K., MICHALAK, A., MUSIK, I. & BIALA, G. 2015. Effects of imperatorin on scopolamine-induced cognitive impairment and oxidative stress in mice. *Psychopharmacology (Berl)*, 232, 931-42.
- CAO, Z., WANG, F., XIU, C., ZHANG, J. & LI, Y. 2017. Hypericum perforatum extract attenuates behavioral, biochemical, and neurochemical abnormalities in Aluminum chloride-induced Alzheimer's disease rats. *Biomed Pharmacother*, 91, 931-937.
- CHAVALI, V. D., AGARWAL, M., VYAS, V. K. & SAXENA, B. 2020. Neuroprotective Effects of Ethyl Pyruvate against Aluminum Chloride-Induced Alzheimer's Disease in Rats via Inhibiting Toll-Like Receptor 4. *J Mol Neurosci*, 70, 836-850.
- CHIROMA, S. M., BAHARULDIN, M. T. H., MAT TAIB, C. N., AMOM, Z., JAGADEESAN, S., ILHAM ADENAN, M., MAHDI, O. & MOKLAS, M. A. M. 2019. Centella asiatica protects d-galactose/AlCl<sub>3</sub> mediated Alzheimer's disease-like rats via PP2A/GSK-3 $\beta$  signaling pathway in their Hippocampus. *International journal of molecular sciences*, 20, 1871.
- COLOMINA, M. T. & PERIS-SAMPEDRO, F. 2017. Aluminum and Alzheimer's Disease. *Adv Neurobiol*, 18, 183-197.
- F EWIDA, S. & A MANSOUR, M. 2015. the antioxidant metabolic protective effect of exercise on d-galactose induced spatial memory impairment; possible role of hippocampal astrocytes. *Al-Azhar Medical Journal*, 44, 295-316.
- FARZAMPOUR, S., MAJDI, A. & SADIGH-ETEGHAD, S. 2016. Intranasal insulin treatment improves memory and learning in a rat amyloid-beta model of Alzheimer's disease. *Physiol Int*, 103, 344-353.
- FAWZI, S. F., MENZE, E. T. & TADROS, M. G. 2020. Deferiprone ameliorates memory impairment in Scopolamine-treated rats: The impact of its iron-chelating effect on  $\beta$ -amyloid disposition. *Behav Brain Res*, 378, 112314.
- FERREIRA, M. E. S., DE VASCONCELOS, A. S., DA COSTA VILHENA, T., DA SILVA, T. L., DA SILVA BARBOSA, A., GOMES, A. R. Q., DOLABELA, M. F. & PERCÁRIO, S. 2015. Oxidative stress in Alzheimer's disease: should we keep trying antioxidant therapies? *Cellular and molecular neurobiology*, 35, 595-614.
- FIRDAUS, Z., SINGH, N., PRAJAPATI, S. K., KRISHNAMURTHY, S. & SINGH, T. D. 2022. Centella asiatica prevents D-galactose-Induced cognitive deficits, oxidative stress and neurodegeneration in the adult rat brain. *Drug Chem Toxicol*, 45, 1417-1426.
- FUJI, T., INOUE, T. & HASEGAWA, Y. 2018. Nacre extract prevents scopolamine-induced memory deficits in rodents. *Asian Pacific Journal of Tropical Medicine*, 11, 202.
- GAWEL, K., GIBULA, E., MARSZALEK-GRABSKA, M., FILAROWSKA, J. & KOTLINSKA, J. H. 2019. Assessment of spatial learning and memory in the Barnes maze task in rodents-methodological consideration. *Naunyn Schmiedebergs Arch Pharmacol*, 392, 1-18.
- GHANBARI, A., VAFAEI, A. A., ATTARMOGHADDAM, M., BANDEGI, A. & MORADI-KOR, N. 2019. Spirulina microalgae improves memory deficit induced by scopolamine in male pup rats: Role of oxidative stress. *South African Journal of Botany*, 127, 220-225.
- HAFEZ, H. S., GHAREEB, D. A., SALEH, S. R., ABADY, M. M., EL DEMELLAWY, M. A., HUSSIEN, H. & ABDEL-MONEM, N. 2017. Neuroprotective effect of ipriflavone against scopolamine-induced memory impairment in rats. *Psychopharmacology (Berl)*, 234, 3037-3053.
- HANSSON, O., ZETTERBERG, H., BUCHHAVE, P., LONDOS, E., BLENNOW, K. & MINTHON, L. 2006. Association between CSF biomarkers and incipient Alzheimer's disease in patients with mild cognitive impairment: a follow-up study. *Lancet Neurol*, 5, 228-34.
- HARANDI, S., GOLCHIN, L., ANSARI, M., MORADI, A., SHABANI, M. & SHEIBANI, V. 2015. Antiamnesic Effects of Walnuts Consumption on Scopolamine-Induced Memory Impairments in Rats. *Basic Clin Neurosci*, 6, 91-9.
- HEO, Y. M., SHIN, M. S., KIM, S. H., KIM, T. W., BAEK, S. B. & BAEK, S. S. 2014. Treadmill exercise ameliorates disturbance of spatial learning ability in scopolamine-induced amnesia rats. *J Exerc Rehabil*, 10, 155-61.
- IMBIMBO, B. P., LOMBARD, J. & POMARA, N. 2005. Pathophysiology of Alzheimer's disease. *Neuroimaging Clin N Am*, 15, 727-53, ix.



- INAN-EROGLU, E. & AYAZ, A. 2018. Is aluminum exposure a risk factor for neurological disorders? *J Res Med Sci*, 23, 51.
- JUSTIN-THENMOZHI, A., DHIVYA BHARATHI, M., KIRUTHIKA, R., MANIVASAGAM, T., BORAH, A. & ESSA, M. M. 2018. Attenuation of Aluminum Chloride-Induced Neuroinflammation and Caspase Activation Through the AKT/GSK-3 $\beta$  Pathway by Hesperidin in Wistar Rats. *Neurotox Res*, 34, 463-476.
- KANDİŞ, S., ALTINTAŞ, L., KIZILDAĞ, S., SIMSEK, F., ATEŞ, M., ALACAM, H. & UYSAL HARZADIN, N. 2022. Comparison anti-oxidant and neuroprotective effects of extra-virgin olive oil, donepezil and rosmarinic acid on aluminum chloride-induced Alzheimer's in rat models. *ANKARA UNIVERSITESI VETERINER FAKULTESI DERGISI*, 69.
- KARRAN, E. & DE STROOPER, B. 2016. The amyloid cascade hypothesis: are we poised for success or failure? *J Neurochem*, 139 Suppl 2, 237-252.
- KIM, M. H., KIM, S. H. & YANG, W. M. 2014. Mechanisms of action of phytochemicals from medicinal herbs in the treatment of Alzheimer's disease. *Planta Med*, 80, 1249-58.
- KLOTZ, K., WEISTENHÖFER, W., NEFF, F., HARTWIG, A., VAN THRIEL, C. & DREXLER, H. 2017. The Health Effects of Aluminum Exposure. *Dtsch Arztebl Int*, 114, 653-659.
- KNIERIM, J. J. 2015. The hippocampus. *Current Biology*, 25, R1116-R1121.
- KNOPMAN, D. S., AMIEVA, H., PETERSEN, R. C., CHÉTELAT, G., HOLTZMAN, D. M., HYMAN, B. T., NIXON, R. A. & JONES, D. T. 2021. Alzheimer disease. *Nature reviews Disease primers*, 7, 1-21.
- KOU, X., LI, J., LIU, X., YANG, X., FAN, J. & CHEN, N. 2017. Ampelopsin attenuates the atrophy of skeletal muscle from d-gal-induced aging rats through activating AMPK/SIRT1/PGC-1 $\alpha$  signaling cascade. *Biomedicine & Pharmacotherapy*, 90, 311-320.
- KUMAR, A., PRAKASH, A. & DOGRA, S. 2011. Centella asiatica attenuates D-galactose-induced cognitive impairment, oxidative and mitochondrial dysfunction in mice. *International journal of Alzheimer's disease*, 2011.
- KUMAR, A., SIDHU, J., GOYAL, A. & TSAO, J. W. 2018. Alzheimer disease.
- LI, R., HUANG, Z., LUO, J., LUO, H. & WANG, W. 2020. Downregulation of the CB1-Mediated Endocannabinoid Signaling Underlies D-Galactose-Induced Memory Impairment. *Frontiers in Molecular Neuroscience*, 13.
- LIVINGSTON, G., HUNTLEY, J., SOMMERLAD, A., AMES, D., BALLARD, C., BANERJEE, S., BRAYNE, C., BURNS, A., COHEN-MANSFIELD, J. & COOPER, C. 2020. Dementia prevention, intervention, and care: 2020 report of the Lancet Commission. *The Lancet*, 396, 413-446.
- MAT NURI, T. H., HONG, Y. H., MING, L. C., MOHD JOFFRY, S., OTHMAN, M. F. & NEOH, C. F. 2017. Knowledge on Alzheimer's Disease among Public Hospitals and Health Clinics Pharmacists in the State of Selangor, Malaysia. *Front Pharmacol*, 8, 739.
- MAYA, S., PRAKASH, T., MADHU, K. D. & GOLI, D. 2016. Multifaceted effects of aluminium in neurodegenerative diseases: A review. *Biomedicine & Pharmacotherapy*, 83, 746-754.
- NAM, S. M., KIM, J. W., YOO, D. Y., KIM, W., JUNG, H. Y., CHOI, J. H., HWANG, I. K., SEONG, J. K. & YOON, Y. S. 2016. Effects of aluminum on the reduction of neural stem cells, proliferating cells, and differentiating neuroblasts in the dentate gyrus of D-galactose-treated mice via increasing oxidative stress. *J Vet Sci*, 17, 127-36.
- NEWAIRY, A. S., SALAMA, A. F., HUSSIEN, H. M. & YOUSEF, M. I. 2009. Propolis alleviates aluminium-induced lipid peroxidation and biochemical parameters in male rats. *Food Chem Toxicol*, 47, 1093-8.
- OGUNLADE, B., ADELAKUN, S. A. & AGIE, J. A. 2022. Nutritional supplementation of gallic acid ameliorates Alzheimer-type hippocampal neurodegeneration and cognitive impairment induced by aluminum chloride exposure in adult Wistar rats. *Drug Chem Toxicol*, 45, 651-662.
- OHKAWA, H., OHISHI, N. & YAGI, K. 1979. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Analytical biochemistry*, 95, 351-358.
- PADURARIU, M., CIOBICA, A., MAVROUDIS, I., FOTIOU, D. & BALOYANNIS, S. 2012. Hippocampal neuronal loss in the CA1 and CA3 areas of Alzheimer's disease patients. *Psychiatr Danub*, 24, 152-8.
- PATTANASHETTI, L. A., TARANALLI, A. D., PARVATRAO, V., MALABADE, R. H. & KUMAR, D. 2017. Evaluation of neuroprotective effect of quercetin with donepezil in scopolamine-induced amnesia in rats. *Indian Journal of Pharmacology*, 49, 60.
- RAVI, S. K., RAMESH, B. N., MUNDUGARU, R. & VINCENT, B. 2018. Multiple pharmacological activities of Caesalpinia crista against aluminium-induced neurodegeneration in rats: Relevance for Alzheimer's disease. *Environmental toxicology and pharmacology*, 58, 202-211.
- RETINASAMY, T., SHAIKH, M. F., KUMARI, Y. & OTHMAN, I. 2019. Ethanolic Extract of Orthosiphon stamineus Improves Memory in Scopolamine-Induced Amnesia Model. *Front Pharmacol*, 10, 1216.
- SAEED, A., QUSTI, S. Y., ALMARWANI, R. H., JAMBI, E. J., ALSHAMMARI, E. M., GUSTY, N. F. & BALGOON, M. J. 2021. Effects of aluminum chloride and coenzyme Q10 on the molecular structure of lipids and the morphology of the brain hippocampus cells. *RSC Adv*, 11, 29925-29933.
- SAENNO, R., DORNLAORN, O., ANOSRI, T., KAEWNGAM, S., SIRICHOAT, A., ARANAROCHANA, A., PANNANGRONG, W., WIGMORE, P. & WELBAT, J. U. 2022. Caffeic Acid Alleviates Memory and Hippocampal



- Neurogenesis Deficits in Aging Rats Induced by D-Galactose. *Nutrients*, 14.
- SAHRAEI, H., ASKARIPOUR, M., ESMAEILPOUR, K., SHAHSAVARI, F., RAJABI, S. & MORADIKOR, N. 2019. GABA(B) receptor activation ameliorates spatial memory impairments in stress-exposed rats. *Neuropsychiatr Dis Treat*, 15, 1497-1506.
- SHWE, T., PRATCHAYASAKUL, W., CHATTIPAKORN, N. & CHATTIPAKORN, S. C. 2018. Role of D-galactose-induced brain aging and its potential used for therapeutic interventions. *Exp Gerontol*, 101, 13-36.
- SINGH, N. A., BHARDWAJ, V., RAVI, C., RAMESH, N., MANDAL, A. K. A. & KHAN, Z. A. 2018. EGCG Nanoparticles Attenuate Aluminum Chloride Induced Neurobehavioral Deficits, Beta Amyloid and Tau Pathology in a Rat Model of Alzheimer's Disease. *Front Aging Neurosci*, 10, 244.
- TAMIZHAZHAGAN, V. & PUGAZHENDY, K. 2017. Histological methods in life science. *International Journal of Biomedical Materials Research*, 5, 68.
- UGBAJA, R. N., OLANIYAN, T. O., AFOLABI, O. K., ONUNKWOR, B. O., DOSUMU, O. A., AKINHANMI, T. F., TIAMIYU, A. Y. & ALIYU-BANJO, N. O. 2017. Ionoregulatory Disruption and Acetylcholinesterase Activity in Aluminium Toxicity: Effects of Vitamins C and E. *Niger J Physiol Sci*, 32, 207-212.
- UM, Y. H., CHOI, W. H., JUNG, W. S., PARK, Y. H., LEE, C. U. & LIM, H. K. 2017. A Case Report of a 37-Year-Old Alzheimer's Disease Patient with Prominent Striatum Amyloid Retention. *Psychiatry Investig*, 14, 521-524.
- VLISSARA, H., BUCALA, R. & STRIKER, L. 1994. Pathogenic effects of advanced glycosylation: biochemical, biologic, and clinical implications for diabetes and aging. *Lab Invest*, 70, 138-51.
- WANG, C., SHEN, Z., YU, J., YANG, J., MENG, F., JIANG, X. & ZHU, C. 2020. Protective effects of enzyme degradation extract from *Porphyra yezoensis* against oxidative stress and brain injury in D-galactose-induced ageing mice. *Br J Nutr*, 123, 975-986.
- WEI, X., WEI, H., YANG, D., LI, D., YANG, X., HE, M., LIN, E. & WU, B. 2018. Effect of Aluminum Exposure on Glucose Metabolism and Its Mechanism in Rats. *Biol Trace Elem Res*, 186, 450-456.
- WONG-GUERRA, M., JIMÉNEZ-MARTIN, J., PARDO-ANDREU, G. L., FONSECA-FONSECA, L. A., SOUZA, D. O., DE ASSIS, A. M., RAMIREZ-SANCHEZ, J., DEL VALLE, R. M. & NUÑEZ-FIGUEREDO, Y. 2017. Mitochondrial involvement in memory impairment induced by scopolamine in rats. *Neurol Res*, 39, 649-659.
- XU, D. L., XU, M. M. & WANG, D. H. 2019. Effect of temperature on antioxidant defense and innate immunity in Brandt's voles. *Zool Res*, 40, 305-316.
- YAKUBU, O. E., NWODO, O., IMO, C. & OGWONI, H. 2017. Spermatogenic and haematological effects of aqueous and ethanolic extracts of *hymenocardiaacida* stem bark on aluminiuminduced toxicity in male wistar rats. *Insights in Biomedicine*, 2, 1-5.
- YANG, M. H., CHEN, S. C., LIN, Y. F., LEE, Y. C., HUANG, M. Y., CHEN, K. C., WU, H. Y., LIN, P. C., GOZES, I. & TYAN, Y. C. 2019. Reduction of aluminum ion neurotoxicity through a small peptide application - NAP treatment of Alzheimer's disease. *J Food Drug Anal*, 27, 551-564.
- YIN, Y. Y., LIU, H., CONG, X. B., LIU, Z., WANG, Q., WANG, J. Z. & ZHU, L. Q. 2010. Acetyl-L-carnitine attenuates okadaic acid induced tau hyperphosphorylation and spatial memory impairment in rats. *J Alzheimers Dis*, 19, 735-46.
- ZHANG, J., TUCKER, L. D., DONGYAN, LU, Y., YANG, L., WU, C., LI, Y. & ZHANG, Q. 2018. Tert-butylhydroquinone post-treatment attenuates neonatal hypoxic-ischemic brain damage in rats. *Neurochem Int*, 116, 1-12.